

Chemical Constituents and Anti-platelet Aggregation Activity from the Root of *Peucedanum formosanum*

YU-CHANG CHEN^{1,2}, PENG-YIN CHEN¹, CHIN-CHUNG WU¹, IAN-LIH TSAI¹ AND IH-SHENG CHEN^{1*}

¹ Graduate Institute of Natural Products, College of Pharmacy, Kaohsiung Medical University, Kaohsiung City 807, Taiwan (R.O.C.)

² School of Chinese Medicine Resources, College of Pharmacy, China Medical University, Taichung City 404, Taiwan (R.O.C.)

(Received: July 13, 2007; Accepted: October 8, 2007)

ABSTRACT

Analysis of the root extract of *Peucedanum formosanum* (Taiwan Qian-Hu) led to the isolation of 32 known compounds. The structures of these isolates were determined by spectral data. Some of them displayed strong anti-platelet aggregation activities. Analysis showed that most of the constituents found in *P. formosanum* were the same as those found in *P. praeruptorum* (Bai-Hua Qian-Hu), in that many isolates of both plants' roots belong to seselin-type dihydropyranocoumarins and psoralen-type furanocoumarins.

Key words: *Peucedanum formosanum*, Umbelliferae, root, Qian-Hu, furanocoumarin, dihydropyranocoumarin, anti-platelet aggregation

INTRODUCTION

Peucedanum formosanum Hay. (Umbelliferae) is an endemic perennial herb in Taiwan and distributed at medium to high altitudes through the island⁽¹⁾. Its root has been used as folk medicine to treat coughs, fever, headache and excessive sputum caused by colds. In this regard it resembles the traditional Chinese medicine Qian-Hu, which is derived from the roots of *P. praeruptorum* (Bai-Hua Qian-Hu) and *Angelica decursiva* (*P. decursivum*, *Porphyroscias decursiva*; Zi-Hua Qian-Hu). Two new compounds, peuformosin and (+)-anomalin, have been isolated by means of ether extraction from the root of *P. formosanum*^(2,3). The methanolic extract of the root exerted anti-platelet aggregation activity in preliminary screening and subsequent investigation, which led to the isolation of 32 known compounds. In this paper, we report the isolation, the anti-platelet aggregation activities of these isolates, and the comparisons of the constituents of several *Peucedanum* species used in folk or traditional Chinese medicine.

MATERIALS AND METHODS

I. General

All melting points were determined on a YANACO

* Author for correspondence. Tel: +886-7-3121101 ext. 2191; Fax: +886-7-3210683; E-mail: m635013@kmu.edu.tw

micro-melting point apparatus and were uncorrected. ¹H-NMR (400 MHz) and ¹³C-NMR (100 MHz) spectra were taken on a Varian Unity plus-400 and Varian Mercury plus-400. Chemical shifts were given in δ with TMS as an internal standard. EI-mass spectra were performed on a VG Biotech Quattro 5002 using a direct inlet system. HR-mass spectra were recorded on a JEOL JMX-HX 110 spectrometer. UV spectra were determined on a Hitachi U-2000 double beam spectrophotometer in methanol (MeOH) solution. IR spectra were recorded on a Perkin Elmer system 2000 FT-IR (KBr or neat) spectrophotometer. Optical rotation was measured on a JASCO P-1020 polarimeter. Column chromatography (CC) was carried out on silica gel (Merck, 70-230 and 230-400 mesh) or Sephadex LH-20 gel (Pharmacia, Fine Chemicals AB, Uppsala). Prep. TLC was run on silica gel plates (Merck, 60 F-254). The HPLC system is consisted of a Hitachi L-7100 pump, a Biscoff RI detector and a silica gel column (LiChroCART[®] 250-10, Merck).

II. Plant

The roots of *P. formosanum* were collected from Wutai, Pingtung County, Taiwan, in Aug. 2003 and identified by Dr. Ih-Sheng Chen, College of Pharmacy, Kaohsiung Medical University. A voucher specimen (Chen 6145) was deposited in the Herbarium of the School of Pharmacy, Kaohsiung Medical University, Kaohsiung, Taiwan, R.O.C.

III. Extraction and Isolation

Dried root (3.4 kg) of *P. formosanum* was sliced and extracted four times with cold MeOH. The precipitate filtered from the concentrated MeOH solution was washed by ethyl acetate (EtOAc), then recrystallized from EtOH to obtain D-mannitol (**1**, 80 g). The filtrate was removed from the solvent in vacuum and partitioned into CHCl₃ soluble fraction (180 g), *n*-BuOH soluble fraction (40 g), and H₂O soluble fraction (360 g). The CHCl₃-soluble fraction (180 g) was chromatographed over silica gel (2.0 kg), eluted with CHCl₃ and gradually enriched with MeOH to give 12 fractions (frs. 1–12).

Fr. 3 (18.5 g; CHCl₃–MeOH, 49:1) was washed by *n*-hexane to get the crystalline mass (11.3 g) which was silica gel CC (330 g) eluted with *n*-hexane/acetone (20:1) with gradually increasing polarity to afford bergapten (**2**, 33.5 mg), isoimperatorin (**3**, 12.2 mg), (–)-deltoidin (**4**, 23.5 mg), and xanthotoxin (**5**, 10.4 mg), a mixture (75.8 mg) of β-sitosterol (**6**) and stigmaterol (**7**), a mixture (23.5 mg) of (+)-praeruptorin E (**8**) and (+)-hyuganin A (**9**), and (–)-*cis*-3'-isovaleryl-4'-seneciolykhellactone (**10**, 4.5 mg). The washings (4.5 g) of fr. 3 were silica gel CC (135 g) and eluted with *n*-hexane–acetone (5:1) to produce panaxynol (**11**, 156 mg).

Fr. 4 (40.1 g; CHCl₃/MeOH, 97:3) was washed with *n*-hexane to obtain the crystalline mass (15.2 g). Part (1.0 g) of the crystalline mass was subjected to preparative HPLC (*n*-hexane/EtOAc, 4:1, flow rate = 3 mL/min) to produce (–)-isosamidin (**12**, 18.5 mg), (+)-peuformosin (**13**, 141 mg), (+)-anomalin (**14**, 86.2 mg), and (+)-*cis*-3'-acetoxy-4'-(2-methylbutyroyloxy)-3',4'-dihydroseselin (**15**, 7.8 mg). The washings (20 g) of fr. 4 were silica gel CC (600 g) eluted with *n*-hexane/EtOAc (10:1), with gradually increasing polarity to obtain a mixture (3.4 mg) of *cis*-3'-hydroxy-4'-isovaleryloxy-3',4'-dihydroseselin (**16**) and laserpitin (**17**), (–)-*cis*-3'-hydroxy-4'-(2-methylbutyryloxy)-3',4'-dihydroseselin (**18**, 6.8 mg), and (+)-marmesin (**19**, 8.9 mg).

Part (5.0 g) of fr. 5 (33.4 g; CHCl₃/MeOH, 24:1) was silica gel CC (150 g) eluted with *n*-hexane/CH₂Cl₂ (5:1), with gradually increasing polarity with CH₂Cl₂/EtOAc (10:1) to produce umbelliferone (**20**, 4.6 mg), isoscopolin (**21**, 1.6 mg), *p*-hydroxyphenethyl ferulate (**22**, 8.9 mg), faltarindiol (**23**, 32.6 mg), and psoralen (**24**, 1.5 mg).

Fr. 7 (3.4 g; CHCl₃/MeOH, 96:4) was silica gel CC (150 g) eluted with CHCl₃/MeOH (10:1), with gradually increasing polarity to afford (+)-lomatatin (**25**, 3.2 mg), isofraxidin (**26**, 1.8 mg), and (–)-*cis*-khellactone (**27**, 3.8 mg).

Fr. 8 (2.6 g; CHCl₃/MeOH, 9:1) was chromatographed on Sephadex LH-20 eluted with MeOH to obtain 1-*O*-hexadecanoyl glycerol (**28**, 2.5 mg), (+)-3'-hydroxymarmesin (**29**, 4.7 mg) and (+)-rutaretin (**30**, 28.3 mg).

Fr. 9 (0.8 g; CHCl₃/MeOH, 4:1) was chromatographed on Sephadex LH-20 eluted with MeOH to afford (+)-oxypeucedanin hydrate (**31**, 1.7 mg) and (+)-dorsteniol (**32**, 1.8 mg).

IV. Isolates

D-Mannitol (**1**): colorless needles, m.p. 166–168°C (EtOH), $[\alpha]_D^{24}$: +45.6° (*c* 1.55, pyridine).

Bergapten (**2**): colorless needles, m.p. 192–194°C (Et₂O), EI-MS *m/z* (%): 216 ($[M]^+$, 100), 201 (31), 173 (57), 145 (20). IR ν_{\max}^{KBr} cm⁻¹: 1731(C=O). UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 222 (4.58), 249 (4.46), 259 (4.42), 267 (4.46), 310 (4.38). ¹H-NMR (CDCl₃, 200 MHz): δ 4.26 (3H, *s*, OMe-5), 6.26 (1H, *d*, *J* = 9.8 Hz, H-3), 7.01 (1H, *br d*, *J* = 2.4 Hz, H-3'), 7.12 (1H, *br s*, H-8), 7.59 (1H, *d*, *J* = 2.4 Hz, H-2'), 8.14 (1H, *d*, *J* = 9.8 Hz, H-4).

Isoimperatorin (**3**): light yellow needles, m.p. 108–110°C (Et₂O), EI-MS *m/z* (%): 270 ($[M]^+$, 0.2), 202 (100), 174 (19), 69 (70). IR ν_{\max}^{KBr} cm⁻¹: 1728 (C=O). UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 222 (4.73), 250 (4.63), 259 (4.58), 267 (4.57), 308 (4.53). ¹H-NMR (CDCl₃, 200 MHz): δ 1.70 (3H, *s*, H-4''), 1.80 (3H, *s*, H-5''), 4.92 (2H, *d*, *J* = 7.4 Hz, H-1''), 5.53 (1H, *br t*, *J* = 7.4 Hz, H-2''), 6.27 (1H, *d*, *J* = 10.0 Hz, H-3), 6.95 (1H, *br d*, *J* = 2.2 Hz, H-3'), 7.15 (1H, *br s*, H-8), 7.59 (1H, *d*, *J* = 2.2 Hz, H-2'), 8.16 (1H, *d*, *J* = 10.0 Hz, H-4).

(–)-Deltoidin (**4**): colorless prisms, m.p. 86–88°C (*n*-hexane), $[\alpha]_D^{25}$: –70.4° (*c* 0.08, CHCl₃), EI-MS *m/z* (%): 328 ($[M]^+$, 0.1), 228 (28), 214 (15), 213 (100), 187 (13), 83 (43). IR ν_{\max}^{KBr} cm⁻¹: 1708 (C=O). UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 222 (4.78), 258 (4.15), 333 (4.17). ¹H-NMR (CDCl₃, 200 MHz): δ 1.60 (3H, *s*, Me-4'), 1.62 (3H, *s*, Me-4'), 1.67 (3H, *s*, H-5''), 1.88 (3H, *br d*, *J* = 6.0 Hz, H-4''), 3.26 (2H, *d*, *J* = 8.4 Hz, H-3'), 5.06 (1H, *t*, *J* = 8.4 Hz, H-2'), 5.98 (1H, *br q*, *J* = 6.0 Hz, H-3''), 6.21 (1H, *d*, *J* = 9.4 Hz, H-3), 6.73 (1H, *s*, H-8), 7.21 (1H, *br s*, H-5), 7.59 (1H, *d*, *J* = 9.4 Hz, H-4).

Xanthotoxin (**5**): colorless needles, m.p. 144–146°C (MeOH), EI-MS *m/z* (%): 216 ($[M]^+$, 100), 201 (23), 173 (46), 145 (21), 89 (26). IR ν_{\max}^{KBr} cm⁻¹: 1713 (C=O). UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 218 (4.54), 248 (4.52), 300 (4.24). ¹H-NMR (CDCl₃, 200 MHz): δ 4.30 (3H, *s*, OMe-8), 6.38 (1H, *d*, *J* = 9.6 Hz, H-3), 6.82 (1H, *d*, *J* = 2.2 Hz, H-3'), 7.36 (1H, *s*, H-5), 7.69 (1H, *d*, *J* = 2.2 Hz, H-2'). 7.77 (1H, *d*, *J* = 9.6 Hz, H-4).

β-Sitosterol (**6**) & stigmaterol (**7**): colorless needles, m.p. 142–144°C (MeOH), $[\alpha]_D^{25}$: –54.3° (*c* 0.08, CHCl₃).

(+)-Praeruptorin E (**8**) & (+)-hyuganin A (**9**): colorless prisms, m.p. 127–130°C (*n*-hexane), $[\alpha]_D^{25}$: +36.18° (*c* 2.78, CHCl₃), GC-EI-MS *m/z* (%): **8**: 428 ($[M]^+$, 0.2), 328 (9), 313 (2), 244 (24), 229 (100), 85 (1.8), 83 (1.6); **9**: 428 ($[M]^+$, 0.2), 328 (10), 313 (12), 244 (12), 229 (100), 85 (3), 83 (6). IR ν_{\max}^{KBr} cm⁻¹: 1735 (C=O). UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 220 (4.94), 245 (4.44), 255 sh (4.31), 300 sh (4.67), 321 (4.85). ¹H-NMR (CDCl₃, 200 MHz): **8**: δ 0.94 (3H, *d*, *J* = 6.2 Hz, H-4'''), 0.95 (3H, *d*, *J* = 6.6 Hz, H-5'''), 1.44 (3H, *s*, Me-2'), 1.47 (3H, *s*, Me-2'), 1.88 (3H, *br q*, *J* = 1.2 Hz, H-5''), 1.96 (3H, *br dq*, *J* = 7.2, 1.2 Hz, H-4''), 2.0–2.4 (3H, *m*, H-2''', 3'''), 5.37 (1H, *d*, *J* = 5.0 Hz, H-3'), 6.12 (1H, *br q*, *J* = 7.2 Hz, H-3''), 6.21 (1H, *d*, *J* = 9.4 Hz, H-3), 6.61 (1H, *d*, *J* = 5.0 Hz, H-4'), 6.80 (1H, *d*, *J* = 8.6 Hz, H-6), 7.35 (1H, *d*, *J* = 8.6 Hz, H-5), 7.60 (1H, *d*, *J* = 9.4 Hz, H-4); **9**: δ 0.89 (3H, *t*, *J* = 7.4 Hz, H-4'''), 1.17 (3H, *d*, *J* =

6.8 Hz, H-5'''), 1.44 (3H, *s*, Me-2'), 1.47 (3H, *s*, Me-2'), 1.56–1.80 (2H, *m*, H-2'''), 5.37 (1H, *d*, $J = 5.0$ Hz, H-3'), 6.21 (1H, *d*, $J = 9.4$ Hz, H-3), 6.59 (1H, *d*, $J = 5.0$ Hz, H-4'), 6.80 (1H, *d*, $J = 8.6$ Hz, H-6), 7.35 (1H, *d*, $J = 8.6$ Hz, H-5), 7.60 (1H, *d*, $J = 9.4$ Hz, H-4).

(–)-*cis*-3'-Isovaleryl-4'-seneciolykhellactone (**10**): colorless oil, $[\alpha]_D^{25}$: -13.3° (*c* 0.08, CHCl₃), EI-MS *m/z* (%): 428 ([M]⁺, 0.1), 326 (5), 311 (10), 261 (5), 244 (8), 229 (39), 231(4.4), 189 (3), 83 (100). IR ν_{\max}^{KBr} cm⁻¹: 1731 (C=O). UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 217 (4.82), 255 sh (4.09), 300 sh (4.32), 323 (3.50). ¹H-NMR (CDCl₃, 200 MHz): δ 0.95 (3H, *d*, $J = 6.2$ Hz, H-4''), 0.97 (3H, *d*, $J = 6.2$ Hz, H-5''), 1.41 (3H, *s*, Me-2'), 1.45 (3H, *s*, Me-2'), 1.89 (3H, *s*, H-4'''), 2.23 (3H, *s*, H-5'''), 1.90–2.20 (3H, *m*, H-2'', 3''), 5.33 (1H, *d*, $J = 4.8$ Hz, H-3'), 5.63 (1H, *br s*, H-2'''), 6.21 (1H, *d*, $J = 9.6$ Hz, H-3), 6.59 (1H, *d*, $J = 4.8$ Hz, H-4'), 6.79 (1H, *d*, $J = 8.8$ Hz, H-6), 7.33 (1H, *d*, $J = 8.8$ Hz, H-5), 7.58 (1H, *d*, $J = 9.6$ Hz, H-4). ¹³C-NMR (CDCl₃, 100 MHz): δ 20.4 (C-5'''), 22.4 (C-4'', 5''), 22.5 (Me-2'), 25.3 (Me-2', C-3''), 27.5 (C-4'''), 43.1 (C-2''), 59.6 (C-4'), 70.2 (C-3'), 77.5 (C-2'), 107.6 (C-8), 112.5 (C-4a), 113.3 (C-3), 114.3 (C-6), 115.1 (C-2'''), 129.0 (C-5), 143.1 (C-4), 154.0 (C-8a), 156.7 (C-7), 158.2 (C-3'''), 159.9 (C-2), 165.1 (C-1''), 171.9 (C-1''').

Panaxynol (**11**): colorless oil, $[\alpha]_D^{25}$: -20.8° (*c* 0.23, CHCl₃), EI-MS *m/z* (%): 244 ([M]⁺, 3), 243 (9), 202 (13), 159 (47), 145 (35), 141 (42), 131 (53), 129 (65), 128 (50), 117 (68), 115 (94), 91 (100). IR ν_{\max}^{neat} cm⁻¹: 3421 (OH), 2233 (C≡C).

(–)-Isosamidin (**12**): colorless needles, m.p. 120–122°C (*n*-hexane), $[\alpha]_D^{25}$: -71.6° (*c* 0.08, CHCl₃), EI-MS *m/z* (%): 386 ([M]⁺, 0.3), 355 (7), 326 (4), 311 (10), 244 (7), 229 (38), 83 (100). IR ν_{\max}^{KBr} cm⁻¹: 1741 (C=O). UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 222 (4.93), 255 (4.36), 300 sh (4.63), 323 (4.81). ¹H-NMR (CDCl₃, 400 MHz): δ 1.42 (3H, *s*, Me-2'), 1.46 (3H, *s*, Me-2'), 1.89 (3H, *d*, $J = 1.2$ Hz, H-4'''), 2.09 (3H, *s*, H-2''), 2.23 (3H, *d*, $J = 1.2$ Hz, H-5'''), 5.30 (1H, *d*, $J = 4.8$ Hz, H-3'), 5.64 (1H, *q*, $J = 1.2$ Hz, H-2'''), 6.22 (1H, *d*, $J = 9.6$ Hz, H-3), 6.58 (1H, *d*, $J = 4.8$ Hz, H-4'), 6.79 (1H, *d*, $J = 8.8$ Hz, H-6), 7.34 (1H, *d*, $J = 8.8$ Hz, H-5), 7.58 (1H, *d*, $J = 9.6$ Hz, H-4).

(+)-Peuformosin (**13**): colorless needles, m.p. 155–156°C (*n*-hexane), $[\alpha]_D^{25}$: $+48.1^\circ$ (*c* 0.43, CHCl₃), EI-MS *m/z* (%): 426 ([M]⁺, 0.1), 326 (4), 311 (7), 244 (4), 229 (22), 213 (3), 189 (1), 83(100). IR ν_{\max}^{KBr} cm⁻¹: 1747 1725 (C=O). UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 218 (4.88), 255 sh (4.41), 300 sh (4.64), 323 (4.48). ¹H-NMR (CDCl₃, 400 MHz): δ 1.38 (3H, *s*, Me-2'), 1.43 (3H, *s*, Me-2'), 1.80 (3H, *br s*, H-5''), 1.81 (3H, *s*, H-4'''), 1.92 (3H, *d*, $J = 7.0$ Hz, H-4''), 2.12 (3H, *s*, H-5'''), 5.32 (1H, *d*, $J = 5.0$ Hz, H-3'), 5.56 (1H, *br s*, H-2'''), 6.05 (1H, *br q*, $J = 7.0$ Hz, H-3''), 6.13 (1H, *d*, $J = 9.5$ Hz, H-3), 6.58 (1H, *d*, $J = 5.0$ Hz, H-4'), 6.75 (1H, *d*, $J = 8.5$ Hz, H-6), 7.31 (1H, *d*, $J = 8.5$ Hz, H-5), 7.55 (1H, *d*, $J = 9.5$ Hz, H-4). ¹³C-NMR (CDCl₃, 100 MHz): δ 15.6 (C-4''), 20.2 (C-5''), 20.2 (C-5'''), 22.2 (Me-2'), 25.5 (Me-2'), 27.3 (C-4'''), 59.3 (C-4'), 70.3 (C-3'), 77.3 (C-2'), 107.4 (C-8), 112.3 (C-4a), 113.0 (C-3), 114.2 (C-6), 115.0

(C-2'''), 127.0 (C-2''), 129.1 (C-5), 139.3 (C-3''), 143.1 (C-4), 153.8 (C-8a), 156.6 (C-7), 157.7 (C-3'''), 159.7 (C-2), 164.8 (C-1'''), 166.1 (C-1'').

(+)-Anomalin (**14**): colorless needles, m.p. 171–173°C (*n*-hexane), $[\alpha]_D^{25}$: $+32.4^\circ$ (*c* 0.2, CHCl₃), EI-MS *m/z* (%): 426 ([M]⁺, 0.1), 327 (37), 311 (53), 229 (100). IR ν_{\max}^{KBr} cm⁻¹: 1731 (C=O), 1604. UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 209 (4.81), 217 (4.80), 255 (4.36), 300 sh (4.45), 323 (4.47). ¹H-NMR (CDCl₃, 400 MHz): δ 1.45 (3H, *s*, Me-2'), 1.49 (3H, *s*, Me-2'), 1.82 (3H, *br s*, H-5''), 1.85 (3H, *br s*, H-5'''), 1.92–1.98 (6H, *m*, H-4'', 4'''), 5.45 (1H, *d*, $J = 5.0$ Hz, H-3'), 6.01 (1H, *br q*, $J = 7.2$ Hz, H-3''), 6.02 (1H, *br q*, $J = 7.2$ Hz, H-3'''), 6.22 (1H, *d*, $J = 9.4$ Hz, H-3), 6.70 (1H, *d*, $J = 5.0$, H-4'), 6.81 (1H, *d*, $J = 8.8$ Hz, H-6), 7.34 (1H, *d*, $J = 8.8$ Hz, H-5), 7.59 (1H, *d*, $J = 9.4$ Hz, H-4).

(+)-*cis*-3'-Acetoxy-4'-(2-methylbutyryloxy)-3',4'-dihydroseselin (**15**): colorless oil, $[\alpha]_D^{25}$: $+12.6^\circ$ (*c* 0.39, CHCl₃), EI-MS *m/z* (%): 388 ([M]⁺, 0.2), 328 (2), 327 (7), 312 (7), 261 (7), 244 (11), 230 (15), 229 (100). IR ν_{\max}^{neat} cm⁻¹: 1743 (C=O), 1608. UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 208 (4.76), 215 sh (4.65), 245 (4.12), 255 (4.05), 300 sh (4.43), 323 (4.62). ¹H-NMR (CDCl₃, 400 MHz): δ 0.94 (3H, *t*, $J = 7.6$ Hz, H-4'''), 1.21 (3H, *d*, $J = 7.2$ Hz, H-5'''), 1.42 (3H, *s*, Me-2'), 1.45 (3H, *s*, Me-2'), 1.48 (1H, *m*, H-3'''), 1.72 (1H, *m*, H-3'''), 2.10 (3H, *s*, H-2''), 2.41 (1H, *m*, H-2'''), 5.30 (1H, *d*, $J = 5.0$ Hz, H-3'), 6.22 (1H, *d*, $J = 9.4$ Hz, H-3), 6.52 (1H, *d*, $J = 5.0$ Hz, H-4'), 6.79 (1H, *d*, $J = 8.6$ Hz, H-6), 7.36 (1H, *d*, $J = 8.6$ Hz, H-5), 7.59 (1H, *d*, $J = 9.4$ Hz, H-4). ¹³C-NMR (CDCl₃, 100 MHz): δ 11.6 (C-4'''), 16.6 (C-5'''), 20.7 (C-2''), 21.9 (Me-2'), 25.5 (Me-2'), 26.6 (C-3'''), 41.3 (C-2'''), 60.3 (C-4'), 70.7 (C-3'), 77.3 (C-2'), 107.4 (C-8), 112.3 (C-4a), 113.3 (C-3), 114.3 (C-6), 129.3 (C-5), 143.2 (C-4), 154.0 (C-8a), 156.5 (C-7), 159.7 (C-2), 169.8 (C-1''), 175.6 (C-1''').

cis-3'-Hydroxy-4'-isovaleryloxy-3',4'-dihydroseselin (**16**) & laserpitin (**17**): colorless oil, $[\alpha]_D^{25}$: -107.5° (*c* 0.22, CHCl₃), GC-EI-MS *m/z* (%): **16**: 346 ([M]⁺, 8), 328 (8), 312 (9), 244 (50), 229 (100); **17**: 344 ([M]⁺, 8), 326 (8), 310 (22), 244 (50), 229 (100). IR ν_{\max}^{neat} cm⁻¹: 3473 (OH), 1729 (C=O), 1606. UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 205 (4.05), 215 sh (3.80), 324 (3.56). ¹H-NMR (CDCl₃, 200 MHz): **16**: δ 0.97 (3H, *d*, $J = 6.2$ Hz, H-4''), 1.01 (3H, *d*, $J = 6.2$ Hz, H-5''), 1.41 (3H, *s*, Me-2'), 1.45 (3H, *s*, Me-2'), 2.10–2.40 (3H, *m*, H-2'', 3''), 2.88 (1H, *br s*, OH-3'), 4.03 (1H, *d*, $J = 5.0$ Hz, H-3'), 6.23 (1H, *d*, $J = 9.6$ Hz, H-3), 6.42 (1H, *d*, $J = 5.0$ Hz, H-4'), 6.80 (1H, *d*, $J = 8.8$ Hz, H-6), 7.35 (1H, *d*, $J = 8.8$ Hz, H-5), 7.60 (1H, *d*, $J = 9.6$ Hz, H-4); **17**: δ 1.41 (3H, *s*, Me-2'), 1.45 (3H, *s*, Me-2'), 1.88 (3H, *br q*, $J = 1.2$ Hz, H-5''), 1.96 (3H, *br dq*, $J = 7.2$, 1.2 Hz, H-4''), 2.88 (1H, *br s*, OH-3'), 4.08 (1H, *d*, $J = 5.0$ Hz, H-3'), 6.12 (1H, *br q*, $J = 7.2$ Hz, H-3''), 6.23 (1H, *d*, $J = 9.6$ Hz, H-3), 6.49 (1H, *d*, $J = 5.0$ Hz, H-4'), 6.80 (1H, *d*, $J = 8.8$ Hz, H-6), 7.35 (1H, *d*, $J = 8.8$ Hz, H-5), 7.60 (1H, *d*, $J = 9.6$ Hz, H-4).

(–)-*cis*-3'-Hydroxy-4'-(2-methylbutyryloxy)-3',4'-dihydroseselin (**18**): colorless oil, $[\alpha]_D^{25}$: -80.5° (*c* 0.34, CHCl₃), EI-MS *m/z* (%): 346 ([M]⁺, 4), 328 (6), 313 (7), 244 (20), 229 (100). IR ν_{\max}^{neat} cm⁻¹: 3472 (OH), 1730 (C=O),

1607. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 206 (4.48), 215 sh (4.16), 246 (3.56), 256 (3.50), 325 (4.12). $^1\text{H-NMR}$ (CDCl_3 , 200 MHz): δ 0.94 (3H, *t*, $J = 7.4$ Hz, H-4''), 1.24 (3H, *d*, $J = 6.8$ Hz, H-5''), 1.42 (3H, *s*, Me-2'), 1.49 (3H, *s*, Me-2'), 1.50 (1H, *m*, H-3''), 1.75 (1H, *m*, H-3''), 2.49 (1H, *m*, H-2''), 2.87 (1H, *br s*, OH-3'), 4.05 (1H, *d*, $J = 5.2$ Hz, H-3'), 6.24 (1H, *d*, $J = 9.6$ Hz, H-3), 6.39 (1H, *d*, $J = 5.2$ Hz, H-4'), 6.79 (1H, *d*, $J = 8.8$ Hz, H-6), 7.36 (1H, *d*, $J = 8.8$ Hz, H-5), 7.61 (1H, *d*, $J = 9.6$ Hz, H-4).

(+)-Marmesin (**19**): colorless prisms, m.p. 187–188°C (*n*-hexane/ CHCl_3), $[\alpha]_{\text{D}}^{25}$: +26.2° (*c* 1.1, CHCl_3), EI-MS *m/z* (%): 246 ($[\text{M}]^+$, 36), 213 (31), 188 (65), 187 (100), 160 (26), 131 (16). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3447 (OH), 1703 (C=O), 1625, 1566. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 207 (4.50), 225 (4.24), 248 (3.81), 258 (3.73), 335 (4.45). $^1\text{H-NMR}$ (CDCl_3 , 200 MHz): δ 1.23 (3H, *s*, Me-4'), 1.37 (3H, *s*, Me-4'), 1.80 (1H, *br s*, OH-4'), 3.22 (2H, *d*, $J = 8.4$ Hz, H-3'), 4.73 (1H, *t*, $J = 8.4$ Hz, H-2'), 6.20 (1H, *d*, $J = 9.6$ Hz, H-3), 6.74 (1H, *s*, H-8), 7.21 (1H, *s*, H-5), 7.59 (1H, *d*, $J = 9.6$ Hz, H-4).

Umbelliferone (**20**): yellow prisms, m.p. 224–226°C (Et_2O /acetone), EI-MS *m/z* (%): 162 ($[\text{M}]^+$, 100), 135 (55), 134 (63), 106 (13), 105 (16), 78 (29). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3161 (OH), 1709 (C=O). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 207 (4.27), 325 (4.31). UV $\lambda_{\text{max}}^{\text{MeOH+KOH}}$ nm (log ϵ): 210 (4.39), 230 (4.09), 371 (4.40). $^1\text{H-NMR}$ (acetone-*d*₆, 200 MHz): δ 6.16 (1H, *d*, $J = 9.4$ Hz, H-3), 6.74 (1H, *d*, $J = 2.2$ Hz, H-8), 6.83 (1H, *dd*, $J = 8.8, 2.2$ Hz, H-6), 7.50 (1H, *d*, $J = 8.8$ Hz, H-5), 7.85 (1H, *d*, $J = 9.4$ Hz, H-4).

Isoscopoletin (**21**): yellow prisms, m.p. 184–186°C (Et_2O /acetone), EI-MS *m/z* (%): 192 ($[\text{M}]^+$, 100), 188 (25), 177 (42), 164 (34), 149 (43), 121 (23), 79 (7), 69 (12). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3442 (OH), 1707 (C=O), 1609, 1567, 1293. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 207 (4.02), 227 (3.75), 255 sh (3.23), 293 (3.30), 338 (3.56). UV $\lambda_{\text{max}}^{\text{MeOH+KOH}}$ nm (log ϵ): 208 (4.09), 244 (3.61), 278 (3.20), 391 (3.66). $^1\text{H-NMR}$ (CDCl_3 , 200 MHz): δ 3.95 (3H, *s*, OMe-7), 6.14 (1H, *br s*, OH-6, D_2O exchangeable), 6.26 (1H, *d*, $J = 9.6$ Hz, H-3), 6.85 (1H, *s*, H-8), 6.92 (1H, *s*, H-5), 7.59 (1H, *d*, $J = 9.6$ Hz, H-4).

p-Hydroxyphenethyl ferulate (**22**): colorless oil, EI-MS *m/z* (%): 314 ($[\text{M}]^+$, 0.5), 194 (100), 177 (12), 145 (18), 120 (36). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3380 (OH), 1693 (C=O), 1595, 1515. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 220 (4.66), 300 sh (4.51), 325 (4.67). UV $\lambda_{\text{max}}^{\text{MeOH+KOH}}$ nm (log ϵ): 210 (4.62), 225 (4.58), 311 (4.20), 378 (4.74). $^1\text{H-NMR}$ (CDCl_3 , 400 MHz): δ 2.94 (2H, *t*, $J = 7.2$ Hz, H-7'), 3.93 (3H, *s*, OMe-3), 4.37 (2H, *t*, $J = 7.2$ Hz, H-8'), 4.95 (1H, *br s*, OH-4, D_2O exchangeable), 5.87 (1H, *s*, OH-4', D_2O exchangeable), 6.27 (1H, *d*, $J = 16.0$ Hz, H-8), 6.78 (2H, *d*, $J = 8.8$ Hz, H-3', 5'), 6.91 (1H, *d*, $J = 8.0$ Hz, H-6), 7.01 (1H, *d*, $J = 2.0$ Hz, H-2), 7.06 (1H, *dd*, $J = 8.0, 2.0$ Hz, H-5), 7.12 (2H, *d*, $J = 8.8$ Hz, H-2', 6'), 7.59 (1H, *d*, $J = 16.0$ Hz, H-7). $^{13}\text{C-NMR}$ (CDCl_3 , 100 MHz): δ 167.3 (C-9), 154.3 (C-4'), 147.9 (C-4), 146.7 (C-3), 144.9 (C-7), 130.0 (C-2', 6'), 129.9 (C-1'), 127.0 (C-1), 123.1 (C-6), 115.3 (C-3', 5', 8), 114.6 (C-5), 109.3 (C-2), 65.1 (C-1''), 55.9 (OMe-3), 34.3 (C-2'').

Falcarindiol (**23**): colorless oil, $[\alpha]_{\text{D}}^{25}$: +95.2° (*c* 0.12, CHCl_3), EI-MS *m/z* (%): 260 ($[\text{M}]^+$, 4), 242 (6), 229 (35), 157 (27), 129 (87), 128 (100), 115 (53), 91 (57). IR $\nu_{\text{max}}^{\text{neat}}$ cm^{-1} : 3380 (OH), 2231, 2146 (C=C). $^1\text{H-NMR}$ (CDCl_3 , 400 MHz): δ 0.88 (3H, *t*, $J = 6.8$ Hz, H-17), 1.25–1.40 (10H, *br s*, H-12–16), 2.11 (2H, *q*, $J = 7.2$ Hz, H-11), 4.93 (1H, *br d*, $J = 5.2$ Hz, H-3), 5.20 (1H, *d*, $J = 8.2$ Hz, H-8), 5.26 (1H, *d*, $J = 10.4$ Hz, Ha-1), 5.47 (1H, *d*, $J = 16.8$ Hz, Hb-1), 5.51 (1H, *dd*, $J = 10.0, 8.2$ Hz, H-9), 5.60 (1H, *dt*, $J = 10.0, 7.2$ Hz, H-10), 5.93 (1H, *ddd*, $J = 16.8, 10.4, 5.2$ Hz, H-2). $^{13}\text{C-NMR}$ (CDCl_3 , 100 MHz): δ 14.1 (C-17), 22.6 (C-16), 27.7 (C-11), 29.0 (C-12), 29.1 (C-13), 29.2 (C-14), 31.7 (C-15), 58.6 (C-8), 63.4 (C-3), 68.6 (C-6), 70.2 (C-5), 78.2 (C-4), 79.8 (C-7), 117.3 (C-1), 127.6 (C-9), 134.7 (C-10), 135.8 (C-2).

Psoralen (**24**): yellow needles, m.p. 161–163°C (Et_2O), EI-MS *m/z* (%): 186 ($[\text{M}]^+$, 100), 158 (64), 130 (20), 102 (25). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 1724 (C=O). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 210 (4.18), 240 sh (4.29), 246 (4.31), 290 (3.94), 328 (3.72).

(+)-Lomatin (**25**): colorless needles, m.p. 156–158°C (*n*-hexane), $[\alpha]_{\text{D}}^{25}$: +45.3° (*c* 0.13, CHCl_3), EI-MS *m/z* (%): 246 ($[\text{M}]^+$, 38), 213 (16), 188 (11), 177 (17), 176 (100), 147 (13), 91 (8). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3440 (OH), 1712 (C=O), 1603. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 209 (4.57), 215 sh (4.16), 247 (3.83), 257 (3.80), 327 (4.49). $^1\text{H-NMR}$ (CDCl_3 , 200 MHz): δ 1.35 (3H, *s*, Me-2'), 1.42 (3H, *s*, Me-2'), 2.97 (1H, *dd*, $J = 17.4, 5.2$ Hz, H-4'), 3.16 (1H, *dd*, $J = 17.4, 5.2$ Hz, H-4'), 3.93 (1H, *t*, $J = 5.2$ Hz, H-3'), 6.24 (1H, *d*, $J = 9.6$ Hz, H-3), 6.79 (1H, *d*, $J = 8.4$ Hz, H-6), 7.26 (1H, *d*, $J = 8.4$ Hz, H-5), 7.64 (1H, *d*, $J = 9.6$ Hz, H-4).

Isorafaxidin (**26**): yellow needles, m.p. 144–146°C (*n*-hexane), EI-MS *m/z* (%): 222 ($[\text{M}]^+$, 100), 207 (29), 194 (35), 179 (29), 167 (27), 161 (20), 149 (49). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3442 (OH), 1703 (C=O), 1264. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 209 (4.30), 220 sh (4.08), 339 (3.75). UV $\lambda_{\text{max}}^{\text{MeOH+KOH}}$ nm (log ϵ): 215 (4.27), 398 (3.97). $^1\text{H-NMR}$ (CDCl_3 , 200 MHz): δ 3.95 (3H, *s*, OMe-6), 4.10 (3H, *s*, OMe-8), 6.13 (1H, *br s*, OH-7, D_2O exchangeable), 6.29 (1H, *d*, $J = 9.6$ Hz, H-3), 6.66 (1H, *s*, H-5), 7.60 (1H, *d*, $J = 9.6$ Hz, H-4).

(-)-*cis*-Khellactone (**27**): colorless prisms, m.p. 145–147°C (Et_2O), $[\alpha]_{\text{D}}^{25}$: -17.4° (*c* 0.18, CHCl_3), EI-MS *m/z* (%): 262 ($[\text{M}]^+$, 15), 192 (13), 191 (100), 162 (16), 134 (16), 107 (14). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3418 (OH), 1712 (C=O), 1603. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 219 (4.67), 245 sh (4.12), 257 (4.04), 300 sh (4.46), 326 (4.70). $^1\text{H-NMR}$ (CDCl_3 , 400 MHz): δ 1.39 (3H, *s*, Me-2'), 1.44 (3H, *s*, Me-2'), 3.23 (1H, *br s*, OH, D_2O exchangeable), 3.84 (1H, *d*, $J = 4.8$ Hz, H-3'), 4.19 (1H, *br s*, OH, D_2O exchangeable), 5.18 (1H, *d*, $J = 4.8$ Hz, H-4'), 6.23 (1H, *d*, $J = 9.6$ Hz, H-3), 6.77 (1H, *d*, $J = 8.6$ Hz, H-6), 7.30 (1H, *d*, $J = 8.6$ Hz, H-5), 7.64 (1H, *d*, $J = 9.6$ Hz, H-4).

1-*O*-Hexadecanoyl glycerol (**28**): amorphous solid, m.p. 66–68°C (CHCl_3), $[\alpha]_{\text{D}}^{25}$: -12.4° (*c* 0.01, CHCl_3), FAB-MS *m/z* (%): 353 ($[\text{M}+\text{Na}]^+$, 19), 313 (12), 239 (14), IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3415 (OH), 1730 (C=O). $^1\text{H-NMR}$ (CDCl_3 , 200 MHz): δ 0.87 (3H, *t*, $J = 6.6$ Hz, H-16'), 1.25–1.30

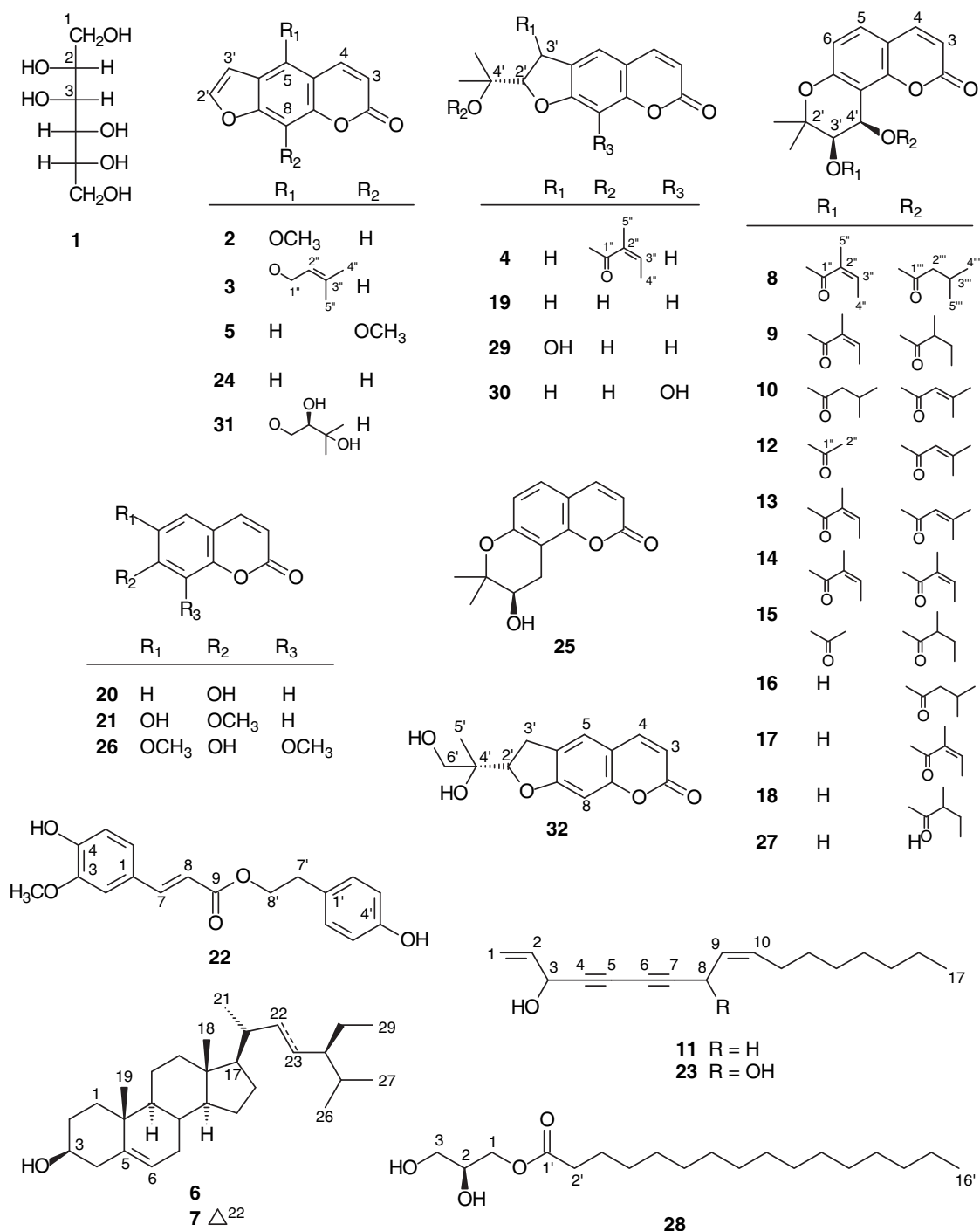


Figure 1. Structures of the root extract of *Peucedanum formosanum* (Taiwan Qian-Hu) led to the isolation of 32 known compounds.

(24H, *br s*, H-4'-15'), 1.62 (2H, *m*, H-3'), 2.35 (2H, *t*, $J = 7.6$ Hz, H-2'), 3.59 (1H, *dd*, $J = 11.4, 5.6$ Hz, H-3), 3.70 (1H, *dd*, $J = 11.4, 4.0$ Hz, H-3), 3.93 (1H, *m*, H-2), 4.13 (1H, *dd*, $J = 11.6, 4.8$ Hz, H-1), 4.22 (1H, *dd*, $J = 11.6, 4.0$ Hz, H-1).

(+)-3'-Hydroxymarmesin (**29**): colorless prisms, m.p. 120–122°C (Et₂O), $[\alpha]_D^{25}$: +36.5° (*c* 0.17, CHCl₃), EI-MS m/z (%): 262 ([M]⁺, 18), 186 (100), 158 (36), 131 (4), 102 (6). IR ν_{\max}^{KBr} cm⁻¹: 3394 (OH), 1725 (C=O), 1628.

UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 205 (4.01), 222 (3.60), 257 sh (2.81), 327 (3.67). ¹H-NMR (CDCl₃, 200 MHz): δ 1.55 (3H, *s*, H-5'), 1.60 (3H, *s*, H-6'), 4.36 (1H, *d*, $J = 6.2$ Hz, H-2'), 5.37 (1H, *br d*, $J = 6.2$ Hz, H-3'), 6.25 (1H, *d*, $J = 9.6$ Hz, H-3), 6.82 (1H, *br s*, H-8), 7.51 (1H, *s*, H-5), 7.65 (1H, *d*, $J = 9.6$ Hz, H-4). ¹³C-NMR (CDCl₃, 50 MHz): δ 25.3 (C-6'), 28.5 (C-5'), 71.8 (C-3'), 73.0 (C-4'), 90.5 (C-2'), 99.0 (C-8), 100.5 (C-4a), 112.9 (C-3), 124.8 (C-5), 128.1 (C-6), 143.6 (C-4), 152.2 (C-7), 156.7 (C-8a), 162.3 (C-2).

(+)-Rutaretin (**30**): yellow prisms, m.p. 179–181°C (*n*-hexane/acetone), $[\alpha]_D^{25}$: +42.2° (*c* 0.09, CHCl₃), EI-MS *m/z* (%): 262 ([M]⁺, 44), 244 (5), 229 (23), 204 (62), 203 (100), 191 (26), 176 (28), 147 (16), 91 (10). IR ν_{\max}^{KBr} cm⁻¹: 3420 (OH), 1700 (C=O), 1618, 1586. UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 213 (4.68), 240 sh (4.13), 266 (3.54), 332 (3.99). UV $\lambda_{\max}^{\text{MeOH+KOH}}$ nm (log ϵ): 219 (4.65), 282 (4.29), 339 (4.23). ¹H-NMR (acetone-*d*₆, 200 MHz): δ 1.23 (3H, *s*, H-5'), 1.29 (3H, *s*, H-6'), 3.24 (1H, *ddd*, *J* = 15.7, 9.2, 1.0 Hz, H-3'), 3.31 (1H, *ddd*, *J* = 15.7, 8.0, 1.0 Hz, H-3'), 3.84 (1H, *br s*, OH-4', D₂O exchangeable), 4.78 (1H, *dd*, *J* = 9.2, 8.0 Hz, H-2'), 6.13 (1H, *d*, *J* = 9.6 Hz, H-3), 6.97 (1H, *t*, *J* = 1.0 Hz, H-5), 7.82 (1H, *d*, *J* = 9.6 Hz, H-4), 8.60 (1H, *br s*, OH-8, D₂O exchangeable). ¹³C-NMR (acetone-*d*₆, 50 MHz): δ 161.4 (C-2), 26.5 (C-5'), 31.8 (C-3'), 72.5 (C-4'), 92.8 (C-2'), 112.9 (C-3), 114.8 (C-4a), 115.6 (C-5), 127.1 (C-6), 130.1 (C-8), 145.2 (C-8a), 146.1 (C-4), 152.5 (C-7).

(+)-Oxypeucedanin hydrate (**31**): colorless needles, m.p. 132–134°C (Et₂O), $[\alpha]_D^{25}$: +26.6° (*c* 0.09, CHCl₃), EI-MS *m/z* (%): 304 ([M]⁺, 23), 259 (30), 202 (91), 167 (100), 147 (29), 137 (47), 59 (45). IR ν_{\max}^{KBr} cm⁻¹: 3146 (OH), 1726 (C=O). UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 220 (4.71), 249 (4.50), 265 sh (4.42), 312 (4.40). ¹H-NMR (CDCl₃, 400 MHz): δ 1.31 (3H, *s*, H-5''), 1.36 (3H, *s*, H-4''), 3.88 (1H, *d*, *J* = 8.0 Hz, H-2''), 4.45 (1H, *dd*, *J* = 10.0, 8.0 Hz, H-1''), 4.53 (1H, *dd*, *J* = 10.0, 3.2 Hz, H-1''), 6.32 (1H, *d*, *J* = 9.6 Hz, H-3), 6.99 (1H, *dd*, *J* = 2.4, 1.2 Hz, H-3'), 7.20 (1H, *d*, *J* = 1.2 Hz, H-8), 7.61 (1H, *d*, *J* = 2.4 Hz, H-2'), 8.18 (1H, *dd*, *J* = 9.6 Hz, H-4).

(+)-Dorsteniol (**32**): colorless prisms, m.p. 135–137°C, $[\alpha]_D^{25}$: +36.8° (*c* 0.09, CHCl₃), EI-MS *m/z* (%): 262 ([M]⁺, 39), 213(35), 188 (73), 187 (100), 160 (20), 131 (24), IR ν_{\max}^{KBr} cm⁻¹: 3415 (OH), 1713 (C=O), 1624. UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ϵ): 210 (4.66), 225 (4.61), 250 sh (4.20), 259 (4.16), 300 sh (4.31), 333 (4.62). ¹H-NMR (CDCl₃, 400 MHz): δ 1.19 (3H, *s*, H-5'), 3.21 (1H, *dd*, *J* = 16.0, 9.6 Hz, H-3'), 3.34 (1H, *dd*, *J* = 16.0, 7.6 Hz, H-3'), 3.48 (1H, *d*, *J* = 11.0 Hz, H-6'), 3.53 (1H, *d*, *J* = 11.0 Hz, H-6'), 4.97 (1H, *dd*, *J* = 9.6, 7.6 Hz, H-2'), 6.14 (1H, *d*, *J* = 9.5 Hz, H-3), 6.64 (1H, *s*, H-8), 7.43 (1H, *s*, H-5), 7.86 (1H, *d*, *J* = 9.5 Hz, H-4).

V. Anti-platelet Aggregation Test

Blood was collected from the marginal vein of a rabbit, anticoagulated with EDTA (6 mM) and centrifuged for 10 min at 90 × *g* at room temperature to obtain platelet-rich plasma (PRP). Platelet suspension was prepared from this EDTA-anticoagulated PRP according to the washing procedures described previously⁽⁴⁾. Platelet numbers were counted by a Coulter counter (Model ZM) and adjusted to 3 × 10⁸ platelets/mL. The platelet pellets were then suspended in Tyrode's solution of the following composition (mM): NaCl (136.8), KCl (2.8), NaHCO₃ (11.9), MgCl₂ (2.1), NaH₂PO₄ (0.33), CaCl₂ (1.0) and glucose (11.2), containing bovine serum albumin (0.35%). Platelet aggregation was measured by turbidimetric method described by O'Brien⁽⁵⁾. The absorbance

of the platelet suspension was taken as 0% aggregation, and that of Tyrode's solution as 100% aggregation. Aggregation was measured by an aggregometer (Chrono-Log Co., Havertown, PA) with consistent stirring at 1200 rpm. All tested compounds were dissolved in dimethyl sulfoxide (DMSO). To eliminate the effect of the solvent on the aggregation, the final concentration of DMSO was fixed at 0.5%, which did not affect the measured aggregation. Aspirin was used as a positive control. Data were analyzed using Student's *t* test.

RESULTS AND DISCUSSION

All of the isolates, including D-mannitol (**1**)⁽⁶⁾, bergapten (**2**)⁽⁷⁾, isoimperatorin (**3**)⁽⁷⁾, (–)-deltoin (**4**)⁽⁷⁾, xanthotoxin (**5**)⁽⁷⁾, β-sitosterol (**6**)⁽⁸⁾, stigmastrol (**7**)⁽⁸⁾, (+)-praeruptorin E (**8**)⁽⁹⁾, (+)-hyuganin A (**9**)⁽¹⁰⁾, (–)-*cis*-3'-isovaleryl-4'-seneciolykhellactone (**10**)⁽¹¹⁾, panaxynol (**11**)⁽¹²⁾, (–)-isosamidin (**12**)⁽⁹⁾, (+)-peuformosin (**13**)⁽³⁾, (+)-anomalin (**14**)⁽⁹⁾, (+)-*cis*-3'-acetoxy-4'-(2-methylbutyryloxy)-3',4'-dihydroseselin (**15**)⁽¹⁰⁾, *cis*-3'-hydroxy-4'-isovaleryloxy-3',4'-dihydroseselin (**16**)⁽⁹⁾, laserpitin (**17**)⁽⁹⁾, (–)-*cis*-3'-hydroxy-4'-(2-methylbutyryloxy)-3',4'-dihydroseselin (**18**)⁽⁹⁾, (+)-marmesin (**19**)⁽⁷⁾, umbelliferone (**20**)⁽¹³⁾, isoscopoletin (**21**)⁽¹⁴⁾, *p*-hydroxyphenethyl ferulate (**22**)⁽¹⁵⁾, faltarindiol (**23**)⁽¹²⁾, psoralen (**24**)⁽⁷⁾, (+)-lomatin (**25**)⁽¹⁶⁾, isofraxidin (**26**)⁽¹⁷⁾, (–)-*cis*-khellactone (**27**)⁽¹³⁾, 1-*O*-hexadecanoyl glycerol (**28**)⁽¹⁸⁾, (+)-3'-hydroxymarmesin (**29**)⁽¹⁹⁾, (+)-rutaretin (**30**)⁽²⁰⁾, (+)-oxypeucedanin hydrate (**31**)⁽⁷⁾, (+)-dorsteniol (**32**)⁽²¹⁾ were readily identified by comparison of physical and spectroscopic data (UV, IR, ¹H-NMR, $[\alpha]_D$, and mass spectrometry data) with values found in the literature. Among these compounds, **11**, **15**, **16**, **18**, **22**, **25**, **26**, **28**, **29**, and **32** were firstly isolated from this genus.

The constituents of three Qian-Hu, Bai-Hua Qian-Hu (*P. praeruptorum*)^(13,22-34), Taiwan Qian-Hu (*P. formosanum*), and Zi-Hua Qian-Hu (*A. decursiva* = *P. decursivum*)⁽³⁵⁻⁴⁷⁾, are compared in Table 1, showing that seselin-type dihydropyranocoumarins (**8–10**, **12–18**, **25**, **27**) and psoralen-type furanocoumarins (**2–5**, **19**, **24**, **28–32**) are two major groups of constituents in *P. formosanum*. These two types of coumarins are also the major compounds in *P. praeruptorum*. The major constituents in *Angelica decursiva* are xanthyletin-type dihydropyranocoumarins and psoralen-type furanocoumarins; the former has not been found in *P. formosanum* and few have been found in *P. praeruptorum*. *P. japonicum*^(2,7,11,48-60), another species of *Peucedanum* in Taiwan⁽¹⁾, also has seselin-type dihydropyranocoumarins and psoralen-type furanocoumarins as its major constituents, and these may be used as a key to the chemotaxonomy of *Peucedanum* and *Angelica*.

Some of the isolates in *P. formosanum* have been reported to exert strong anti-platelet aggregation activities. These include xanthotoxin (**5**)⁽⁷⁾, panaxynol (**11**)⁽⁶¹⁾,

Table 1. The constituents of Bai-Hua Qian-Hu (*Peucedanum praeruptorum*), Taiwan Qian-Hu (*P. formosanum*), and Zi-Hua Qian-Hu (*Angelica decursiva* = *P. decursivum*)

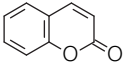
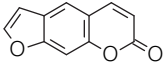
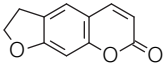
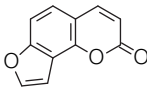
Compound	Bai-Hua Qian-Hu	Taiwan Qian-Hu	Zi-Hua Qian-Hu
			
simple coumarin			
isofraxidin (26)	-	+	-
isoscopoletin (21)	+	+	-
peucedanol	+	-	-
scopoletin	+	-	+
scopolin	+	-	-
umbelliferone (20)	+	+	+
furanocoumarin			
			
psoralen type			
bergapten (2)	-	+	+
decurside I	-	-	+
imperatorin	-	-	+
isoimperatorin (3)	-	+	-
5,8-dimethoxypsoralen	+	-	-
(+)-oxypeucedanin hydrate (31)	-	+	-
psoralen (24)	+	+	-
xanthotoxin (5)	-	+	-
			
dihydropsoresalen type			
decuroside V	-	-	+
decuroside VI	-	-	+
decuroside VII	-	-	+
(-)-deltoidin (4)	-	+	-
(+)-dorsteniol (32)	-	+	-
1- <i>O</i> -hexadecanoyl glycerol (28)	-	+	-
(+)-3'-hydroxymarmesin (29)	-	+	-
isorutarin	+	-	-
(+)-marmesin (19)	-	+	-
nodakenetin	-	-	+
nodakenin	+	-	+
praeroside	+	-	-
(+)-rutaretin (30)	-	+	-
rutarin	+	-	-
			
angelicin type			
angelicin	+	-	-

Table 1. continued

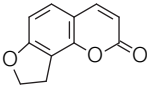
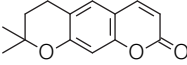
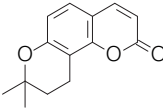
Compound	Bai-Hua Qian-Hu	Taiwan Qian-Hu	Zi-Hua Qian-Hu
			
dihydroangelicin type			
apterin	+	-	-
columbianadin	-	-	+
pyranocoumarin			
			
dihydroxanthyletin type			
3'(<i>S</i>)-acetoxy-4'(<i>R</i>)-isovaleryloxy-3',4'-dihydroxanthyletin	-	-	+
3'(<i>S</i>)-acetoxy-4'(<i>R</i>)-angeloyloxy-3',4'-dihydroxanthyletin	-	-	+
AD-I (3'(<i>S</i>)-angeloyloxy-4'(<i>R,S</i>)-isovaleryloxy-3',4'-dihydroxanthyletin)	-	-	+
andelin (AD-II, 3'(<i>S</i>)-angeloyloxy-4'(<i>R</i>)-senecioyloxy-3',4'-dihydroxanthyletin)	-	-	+
decursidin	-	-	+
decursin	-	-	+
decursinol	+	-	+
decursitin B	-	-	+
decursitin C	-	-	+
decursitin D	-	-	+
decursitin F	-	-	+
Pd-C-I (3'(<i>S</i>)-senecioyloxy-4'(<i>R</i>)-hydroxy-3',4'-dihydroxanthyletin)	+	-	+
Pd-C-II (3'(<i>S</i>)-hydroxy-4'(<i>R</i>)-senecioyloxy-3',4'-dihydroxanthyletin)	-	-	+
Pd-C-III (3'(<i>S</i>)-angeloyloxy-4'(<i>R</i>)-acetoxy-3',4'-dihydroxanthyletin)	-	-	+
qainhucomarin F	+	-	-
			
dihydroseselin type			
(+)- <i>cis</i> -3'-acetoxy-4'-(2-methylbutyroyloxy)-3',4'-dihydroseselin (15)	-	+	-
(+)-anomalin (14, Pd-II)	+	+	-
<i>cis</i> -3',4'-diseneciyl-3',4'-dihydro-seselin	+	-	-

Table 1. continued

Compound	Bai-Hua Qian-Hu	Taiwan Qian-Hu	Zi-Hua Qian-Hu
<i>cis</i> -3'-hydroxy-4'-isovaleryloxy-3',4'-dihydroseselin (16)	-	+	-
(-)- <i>cis</i> -3'-hydroxy-4'-(2-methylbutyryloxy)-3',4'-dihydroseselin (18)	-	+	-
(+)-hyuganin A (9)	-	+	-
(-)-isosamidin (12)	-	+	-
(-)- <i>cis</i> -3'-isovaleryl-4'-senecieryl-khellactone (10)	-	+	-
(-)- <i>cis</i> -khellactone (27)	-	+	-
<i>trans</i> -khellactone	+	-	-
laserpitin (17)	-	+	-
(+)-lomatol (25)	-	+	-
Pd-Ib	+	-	+
peucedanocoumarin I	+	-	-
peucedanocoumarin II	+	-	-
peucedanocoumarin III	+	-	-
(+)-peuformosin (13)	-	+	-
praeroside II	+	-	-
praeroside III	+	-	-
praeroside IV	+	-	-
praeroside V	+	-	-
praeruptorin A	+	-	-
(+)-praeruptorin E (8)	+	+	-
qainhuocoumarin A	+	-	-
qainhuocoumarin B	+	-	-
qainhuocoumarin C	+	-	-
qainhuocoumarin D	+	-	-
qainhuocoumarin E	+	-	-
qainhuocoumarin H	+	-	-
samidin	+	-	-
chromone			
skimmin	+	-	-
steroid			
stigmasterol (7)	+	+	-
β -sitosterol (6)	+	+	+
β -sitosterol- β -D-glucoside	+	-	+
saccharide			
galactitol	+	-	-
D-mannitol (1)	+	+	-
polyacetylene			
falcarindiol (23)	-	+	-
panaxynol (11)	-	+	-
benzenoid			
decursidate	-	-	+
<i>p</i> -hydroxyphenethyl ferulate (22)	-	+	-

(+)-anomalin (**14**)⁽⁶²⁾, psoralen (**24**)⁽⁷⁾ and (-)-*cis*-khellactone (**27**)⁽⁶²⁾. The anti-platelet aggregation effects of the other isolates are shown in Table 2. *p*-Hydroxyphenethyl ferulate (**22**) at 5 μ g/mL showed nearly complete inhibition of platelet aggregation induced by arachidonic acid (AA). (-)-Isosamidin (**12**), (+)-peuformosin (**13**), (+)-*cis*-3'-acetoxy-4'-(2-methylbutyryloxy)-3',4'-dihydroseselin (**15**), *p*-hydroxyphenethyl ferulate (**22**) at 100 μ g/mL showed complete or near complete inhibition of platelet aggregation induced by collagen. (-)-*cis*-3'-Isovaleryl-4'-seneciylkhellactone (**10**), (+)-*cis*-3'-acetoxy-4'-(2-methylbutyryloxy)-3',4'-dihydroseselin (**15**) and *p*-hydroxyphenethyl ferulate (**22**) at 100 μ g/mL abolished platelet-activating factor (PAF)-induced platelet aggregation. Of these compounds, *p*-hydroxyphenethyl ferulate (**22**) showed the strongest antiplatelet aggregation activities, with IC₅₀ values of 5.1, 10.5 and 99.4 μ M for platelet aggregation induced by AA, collagen and PAF, respectively.

ACKNOWLEDGEMENTS

This research was financially supported by the National Science Council of the Republic of China.

REFERENCES

- Kao, M. T. 1993. Umbelliferae. In "Flora of Taiwan". 2nd. ed. Vol. III. pp. 1010-1045. Editorial Committee of the Flora of Taiwan. Taipei, Taiwan.
- Hata, K., Koawa, M., Ikeshiro, Y. and Yen, K. Y. 1968. New coumarins isolated from the root of *Peucedanum formosanum* and *Peucedanum japonicum*. Yakugaku Zasshi 88: 513-520.
- Hata, K., Kozawa, M. and Yen, K. Y. 1966. Constitution of peuformosin, a new coumarin isolated from the root of *Peucedanum formosanum*. Chem. Pharm. Bull. 14: 442-446.
- Teng, C. M., Chen, W. Y., Ko, W. C. and Ouyang, C. 1987. Antiplatelet effect of butylidenephthalide. Biochim. Biophys. Acta 924: 375-382.
- O'Brien, J. R. 1962. Platelet aggregation II. Some results from a new method of study. J. Clin. Path. 15: 452-455.
- Pomilio, A. B., Gonzalez, M. D. and Eceizabarrena, C. C. 1996. 7,8-Dihydroajugasterone C, norhygrine and other constituents of *Nierembergia hippomanica*. Phytochemistry 41: 1393-1398.
- Chen, I. S., Chang, C. T., Sheen, W. S., Teng, C. M., Tsai, I. L., Duh, C. Y. and Ko, F. N. 1996. Coumarins and antiplatelet aggregation constituents from Formosan *Peucedanum japonicum*. Phytochemistry 41: 525-530.
- Kojima, H., Sato, N., Hatano, A. and Ogura, H. 1990. Sterol glucosides from *Prunella vulgaris*. Phytochemistry 29: 2351-2355.

Table 2. Inhibitory effects^a of compounds on the aggregation of washed rabbit platelets induced by thrombin (Thr), arachidonic acid (AA), collagen (Col) and platelet-activating factor (PAF)

Compound	Conc. ($\mu\text{g/mL}$)	Aggregation (%)			
		AA (100 μM)	Col (10 $\mu\text{g/mL}$)	Thr (0.1 U/mL)	PAF (2 ng/mL)
control		84.2 \pm 1.0 (3)	84.8 \pm 2.3 (3)	89.3 \pm 1.6 (3)	87.8 \pm 0.5 (3)
(-)- <i>cis</i> -3'-isovaleryl-4'-seneciylkhel- lactone (10)	100	91.4 \pm 3.2 (3)	32.8 \pm 15.0 (3) ^d	86.3 \pm 1.9 (3)	0.0 \pm 0.0 (3) ^e
	50		49.0 \pm 8.1 (3) ^e		27.0 \pm 14.9 (3) ^d
	20		81.8 \pm 5.0 (3)		58.5 \pm 1.9 (3) ^e
(-)-isosamidin (12)	100	38.0 \pm 17.1 (3) ^e	5.4 \pm 2.8 (3) ^e	87.8 \pm 1.1 (3)	50.3 \pm 15.9 (3) ^e
	50	45.8 \pm 19.3 (3)	30.7 \pm 15.1 (3) ^d		
	20	69.3 \pm 6.6 (3)	68.9 \pm 9.6 (3)		
	10	76.9 \pm 5.4 (3)	77.4 \pm 5.5 (3)		
	5	79.9 \pm 4.3 (3)	83.4 \pm 3.4 (3)		
(+)-peuformosin (13)	100	56.8 \pm 7.7 (3) ^e	14.0 \pm 9.7 (3) ^e	82.1 \pm 2.4 (3) ^e	32.9 \pm 13.3 (3) ^e
(+)- <i>cis</i> -3'-acetoxy-4'-(2-methylbutyroyl- loxy)-3',4'-dihydroseselin (15)	100	53.4 \pm 10.5 (3) ^d	0.0 \pm 0.0 (3) ^e	74.4 \pm 2.8 (3) ^e	0.0 \pm 0.0 (3) ^e
	50	76.1 \pm 2.0 (3) ^e	45.4 \pm 9.3 (3) ^e		22.7 \pm 9.3 (3) ^e
	20		83.4 \pm 1.1 (3) ^e		70.4 \pm 4.9 (3) ^d
(-)- <i>cis</i> -3'-hydroxy-4'-(2-methylbutyryl- loxy)-3',4'-dihydroseselin (18)	100	73.9 \pm 7.0 (3)	66.4 \pm 3.9 (3) ^e	81.0 \pm 3.7 (3)	19.5 \pm 9.0 (3) ^e
	50				66.1 \pm 4.9 (3) ^e
<i>p</i> -hydroxyphenethyl ferulate (22)	100	0.0 \pm 0.0 (3) ^e	0.0 \pm 0.0 (3) ^e	43.8 \pm 8.7 (3) ^e	0.0 \pm 0.0 (3) ^e
	50	0.0 \pm 0.0 (3) ^e	0.0 \pm 0.0 (3) ^e	80.6 \pm 6.7 (3)	9.5 \pm 7.7 (3) ^e
	20	0.0 \pm 0.0 (3) ^e	0.0 \pm 0.0 (3) ^e		74.1 \pm 5.1 (3) ^e
	10	0.0 \pm 0.0 (3) ^e	0.0 \pm 0.0 (3) ^e		
	5	3.1 \pm 2.5 (3) ^e	25.1 \pm 8.9 (3) ^e		
	2	34.5 \pm 17.4 (3) ^e	69.5 \pm 8.7 (3) ^e		
	1	74.7 \pm 3.1 (3) ^d	85.8 \pm 2.8 (3)		
(+)-lomatol (25)	100	81.0 \pm 1.4 (3)	84.7 \pm 0.5 (3) ^e	81.4 \pm 7.6 (3)	73.6 \pm 5.7 (3) ^e
(+)-3'-hydroxymarmesin (29)	100	83.6 \pm 3.1 (3)	90.8 \pm 1.6 (3)	83.4 \pm 3.0 (3)	83.3 \pm 2.6 (3) ^e
(+)-rutaretin (30)	100	64.4 \pm 7.5 (3) ^d	66.6 \pm 3.0 (3) ^e	73.0 \pm 8.3 (3)	86.0 \pm 1.5 (3)
aspirin ^b	100		0.0 \pm 0.0 (3) ^e	81.3 \pm 0.5 (3)	
	50		11.7 \pm 10.1 (4) ^e		
	20		84.3 \pm 0.6 (4) ^e		

^aPlatelets were preincubated with each compound or DMSO (0.5 %, control) at 37°C for 3 min, then the inducer arachidonic acid (AA), collagen, thrombin or PAF was added. Values are presented as mean \pm s.e.m. (n).

^bPositive control.

^cP < 0.05; ^dP < 0.01; ^eP < 0.001 as compared with the respective control.

9. Swager, T. M. and Cardellina II, J. H. 1985. Coumarins from *Musineon divaricatum*. *Phytochemistry* 24: 805-813.

10. Matsuda, H., Murakami, T., Nishida, N., Kageura, T. and Yoshikawa, M. 2000. *Medicinal foodstuff*. XX.

- Vasorelaxant active constituents from the roots of *Angelica furcijuga* Kitagawa: structure of hyuganins A, B, C, and D. *Chem. Pharm. Bull.* 48: 1429-1435.
11. Jong, T. T., Hwang, H. C., Jean M. Y., Wu, T. S. and Teng, C. M. 1992. An antiplatelet aggregation principle and X-ray structural analysis of *cis*-khellactone diester from *Peucedanum japonicum*. *J. Nat. Prod.* 55: 1396-1401.
 12. Fujioka, T., Furumi, K., Fujii, H., Okabe, H., Mihashi, K., Nakano, Y., Matsunaga, H., Katano, M. and Mori, M. 1999. Antiproliferative constituents from Umbelliferae plants. V. A new furanocoumarin and falcariindiol furanocoumarin ethers from the roots of *Angelica japonica*. *Chem. Pharm. Bull.* 47: 96-100.
 13. Kong, L. Y., Li, Y., Min, Z. D., Li, X. and Zhu, T. R. 1996. Coumarins from *Peucedanum praeruptorum*. *Phytochemistry* 41: 1423-1426.
 14. Lee, C. K., Lee, P. H. and Kuo, Y. H. 2001. The chemical constituents from the aril of *Cassia fistula* L. *J. Chin. Chem. Soc.* 48: 1053-1058.
 15. Darwish, F. M. M. and Reinecke, M. G. 2003. Ecdysteroids and other constituents from *Sida spinosa* L. *Phytochemistry* 62: 1179-1184.
 16. Lemmich, J. and Shabana, M. 1984. Coumarin sulphates of *Seseli libanotis*. *Phytochemistry* 23: 863-865.
 17. Panichayupakaranant, P., Noguchi, H., De-Eknamkul, W. and Sankawa, U. 1995. Naphthoquinones and coumarins from *Impatiens balsamina* root cultures. *Phytochemistry* 40: 1141-1144.
 18. Haraldsson, G. G., Gudmundsson, B. O. and Almarsson, O. 1995. The synthesis of homogeneous triglycerides of eicosapentaenoic acid and docosahexaenoic acid by lipase. *Tetrahedron* 51: 941-952.
 19. Lemmich, J., Havelund, S. and Thastrup, O. 1983. Dihydrofurocoumarin glucosides from *Angelica archangelica* and *Angelica silvestris*. *Phytochemistry* 22: 553-555.
 20. Reisch, J. and Voerste, A. A. W. 1994. Natural product chemistry. Part 181. Investigations on the synthesis of dihydropyrano- and dihydrofurano-coumarins by application of catalytic enantioselective *cis*-dihydroxylation. *J. Chem. Soc. Perkin Trans. 1*: 3251-3256.
 21. Ricardo, T. M., Raul, C. G., Norma, F. S. S. and Pedro, J. N. 1998. Isolation, total synthesis, and relative stereochemistry of a dihydrofurocoumarin from *Dorstenia contrajerva*. *J. Nat. Prod.* 61: 1216-1220.
 22. Lu, M., Nicoletti, M., Battinelli, L. and Mazzanti, G. 2001. Isolation of praeruptorins A and B from *Peucedanum praeruptorum* and their general pharmacological evaluation in comparison with extracts of the drug. *Farmaco* 56: 417-420. (C. A. 136: 79275).
 23. Ye, J., Zhang, H. and Yuan, C. 1982. Isolation and identification of coumarin praeruptorin E from the root of the Chinese drug *Peucedanum praeruptorum* Dunn. (Umbelliferae). *Yaoxue Xuebao* 17: 431-434.
 24. Takata, M., Okuyama, T. and Shibata, S. 1988. Studies on coumarins of a Chinese drug "Qian-Hu" VIII. Structures of new coumarin glycoside of Bai-Hua Qian-Hu. *Planta Med.* 54: 323-327.
 25. Okuyama, T., Takata, M. and Shibata, S. 1990. Structures of angular pyranocoumarins of Bai-Hua Qian-Hu, the root of *Peucedanum praeruptorum*. *Planta Med.* 56: 307-311.
 26. Kong, L. Y., Pei, Y. H., Li, X., Wang, S. X., Hou, B. L. and Zhu, T. R. 1993. The isolation and identification of qianhuocoumarin B and C from *Peucedanum praeruptorum*. *Yaoxue Xuebao* 28: 772-776.
 27. Kong, L., Pei, Y., Li, X. and Zhu, T. 1993. New compounds from *Peucedanum praeruptorum*. *Chin. Chem. Lett.* 4: 37-38 (C. A. 121: 31073).
 28. Chang, H. and Li, X. 1999. Coumarins from *Peucedanum praeruptorum* Dunn. *Shenyang Yaoke Daxue Xuebao* 16: 103-106.
 29. Kong, L., Li, Y. and Masatake, N. 2003. A new pyranocoumarin from *Peucedanum praeruptorum*. *Heterocycles* 60: 1915-1919.
 30. Ghen, Z. X., Hung, B. S., She, Q. L. and Zeng, G. F. 1979. Study on the chemical constituents of the Chinese medicinal plant, *Peucedanum praeruptorum* Dunn. Structures of four new coumarins. *Yao Hsueh Hsueh Pao* 14: 486-496.
 31. Okuyama, T. and Shibata, S. 1981. Studies on coumarins of a Chinese drug "Qian-Hu". *Planta Med.* 42: 89-96.
 32. Okuyama, T., Takata, M. and Shibata, S. 1989. Studies on coumarins of a Chinese drug "Qian-Hu" IX. Structures of linear furano- and simple-coumarin glycosides of Bai-Hua Qian-Hu. *Planta Med.* 55: 64-67.
 33. Kong, L. Y., Pei, Y. H., Li, X., Yu, R. M., Min, Z. D. and Zhu, T. R. 1994. Isolation and structure elucidation of baihuaqainhuoside and Pd-C-I from *Peucedanum praeruptorum*. *Yaoxue Xuebao* 29: 276-280.
 34. Kong, L., Pei, Y., Li, X. and Zhu, T. 1993. Chemical constituents of the root of whiteflower hogfennel (*Peucedanum praeruptorum*). *Zhongcaoyao* 24: 401-404.
 35. Kong, L., Li, Y. and Min, Z. 1996. Study on analysis of Pd-Ia in *Peucedanum praeruptorum* and Pd-C-I in *Peucedanum decursivum* by RP-HPLC. *Zhongguo Yaoke Daxue Xuebao* 27: 215-218. (C. A. 125: 323017).
 36. Arima, J. 1929. The constitution of nodakenin, a new glucoside from *Peucedanum decursivum* Maxim. *I. Bull. Chem. Soc. Japan* 4: 113-119.
 37. Kong, L. and Yao, N. H. 2000. Coumarin-glycoside and ferulate from *Peucedanum decursivum*. *Chin. Chem. Lett.* 11: 315-318.
 38. Sakakibara, I., Okuyama, T. and Shibata, S. 1982. Studies on coumarins of a Chinese drug "Qian-Hu". III. Coumarins from "Zi-Hua Qian-Hu". *Planta Med.* 44: 199-103.
 39. Yao, N. and Kong, L. 2001. Chemical constituents of *Peucedanum decursivum*. *Yaoxue Xuebao* 36: 351-355.
 40. Yao, N., Kong, L. and Ling, Y. 1999. New coumarins from *Peucedanum decursivum*. *Chin. Chem. Lett.* 10:

- 477-480.
41. Kong, L., Yao, N. H. and Masatake, N. 2000. Two new xanthyletin-type coumarins from *Peucedanum decursivum*. *Heterocycles* 53: 2019-2025.
 42. Asahara, T., Sakakibara, I., Okuyama, T. and Shibata, S. 1984. Studies on coumarins of a Chinese drug "Qian-Hu" V. Coumarin-glycosides from "Zi-Hua Qian-Hu". *Planta Med.* 50: 488-492.
 43. Sakakibara, I., Okuyama, T. and Shibata, S. 1984. Studies on coumarins of a Chinese drug "Qian-Hu", IV. Coumarins from "Zi-Hua Qian-Hu" (supplement). *Planta Med.* 50: 117-120.
 44. Jung, N. I., Yook, C. S. and Lee, H. K. 1994. Coumarins from the roots of *Angelica decursiva-albiflora*. *Saengyak Hakhoechi* 25: 311-318. (C. A. 122: 101662).
 45. Sano, K., Yosioka, I. and Kitagawa, I. 1973. Stereostructures of decursin, decursidin, and a new coumarin isolated from *Angelica decursiva*. *Chem. Pharm. Bull.* 21: 2095-2097.
 46. Avramenko, L. G., Nikonov, G. K. and Pimenov, M. G. 1970. Andelin, a new dihydropyranocoumarin from *Angelica decursiva* roots. *Khimiya Prirodnykh Soedinenii* 6: 190-194. (C. A. 73: 87814).
 47. Hata, K. and Sano, K. 1969. Coumarins from the root of *Angelica decursiva*. I. Structure of decursin and decursidin. *Yakugaku Zasshi* 89: 549-557. (C. A. 71: 38832).
 48. Yen, K. Y., Lai, H. M. and Chen, W. S. 1966. Chemical constituents of umbelliferous plants of Taiwan. VIII. Coumarins in the roots of *Peucedanum japonicum*. *Taiwan Yaoxue Zazhi* 18: 75-77.
 49. Yamada, Y., Hsu, C. S., Iguchi, K., Suzuki, M., Hsu, H. Y. and Chen, Y. P. 1974. Two new khellactone esters from *Peucedanum japonicum*. *Tetrahedron Lett.* 29: 2513-2516.
 50. Shigematsu, N., Kouno, I. and Kawano, N. 1982. On the isolation of (+)-samidin from the roots of *Peucedanum japonicum* Thunb. *Yakugaku Zasshi* 102: 392-394.
 51. Cisowski, W. 1983. Flavonoid compounds of the herb *Peucedanum japonicum* Thunb. *Pol. J. Chem.* 57: 1283-1286. (C. A. 102: 218357).
 52. Duh, C. Y., Wang, S. K. and Wu, Y. C. 1991. Cytotoxic pyranocoumarins from the aerial parts of *Peucedanum japonicum*. *Phytochemistry* 30: 2812-2814.
 53. Duh, C. Y., Wang, S. K. and Wu, Y. C. 1992. Cytotoxic pyranocoumarins from roots of *Peucedanum japonicum*. *Phytochemistry* 31: 1829-1830.
 54. Ikeshiro, Y., Mase, I. and Tomita, Y. 1992. Dihydropyranocoumarins from roots of *Peucedanum japonicum*. *Phytochemistry* 31: 4303-4306.
 55. Ikeshiro, Y., Mase, I. and Tomita, Y. 1993. Dihydropyranocoumarins from *Peucedanum japonicum*. *Phytochemistry* 33: 1543-1545.
 56. Ikeshiro, Y., Mase, I. and Tomita, Y. 1994. Coumarin glycosides from *Peucedanum japonicum*. *Phytochemistry* 35: 1339-1341.
 57. Lee, G., Park, H. G., Choi, M. L., Kim, Y. H., Park, Y. B., Song, K. S., Cheong, C. and Bae, Y. S. 2000. Falcarindiol, a polyacetylenic compound isolated from *Peucedanum japonicum*, inhibits mammalian DNA topoisomerase I. *J. Microbiol. Biotechnol.* 10: 394-398.
 58. Hisamoto, M., Kikuzaki, H., Ohigashi, H. and Nakatani, N. 2003. Antioxidant compounds from the leaves of *Peucedanum japonicum* Thunb. *J. Agric. Food Chem.* 51: 5255-5261.
 59. Hisamoto, M., Kikuzaki, H. and Nakatani, N. 2004. Constituents of the leaves of *Peucedanum japonicum* Thunb. and their biological activity. *J. Agric. Food Chem.* 52: 445-450.
 60. Lee, S. O., Choi, S. Z., Lee, J. H., Chung, S. H., Park, S. H., Kang, H. C., Yang, E. Y., Cho, H. J. and Lee, K. R. 2004. Antidiabetic coumarin and cyclitol compounds from *Peucedanum japonicum*. *Arch. Pharm. Res.* 27: 1207-1210.
 61. Teng, C. M., Kuo, S. C., Ko, F. N., Lee, J. C., Lee, L. G., Chen, S. C. and Huang, T. F. 1989. Antiplatelet actions of panaxynol and ginsenosides isolated from ginseng. *Biochim. Biophys. Acta* 990: 315-320.
 62. Yoko, A., Toshio, K., Naoki, T. and Seisho, T. 1995. The antagonistic effects of khellactones on platelet-activating factor, histamine, and leukotriene D4. *Chem. Pharm. Bull.* 43: 859-867.